

A-315456: a selective α_{1D} -adrenoceptor antagonist with minimal dopamine D_2 and 5-HT $_{1A}$ receptor affinity

Steven A. Buckner*, Ivan Milicic, Anthony Daza, James J. Lynch III, Teodozjy Kolasa, Masaki Nakane, James P. Sullivan, Jorge D. Brioni

Abbott Laboratories, 100 Abbott Park Road, Abbott Park, IL 60064-6118, USA

Received 28 June 2001; received in revised form 29 October 2001; accepted 6 November 2001

Abstract

In functional assays, A-315456, *N*-[3-(cyclohexylidene-(1*H*-imidazol-4-ylmethyl))phenyl]ethanesulfonamide, behaved as an α_{1D} -adrenoceptor subtype selective antagonist ($pA_2 = 8.34$) in the rat aorta. It was 83-fold less potent at the α_{1B} -adrenoceptor subtype expressed in the rat spleen, and was inactive at the α_{1A} -adrenoceptor subtype expressed in the rat vas deferens. Radioligand binding assays also revealed high affinity ($pK_i = 8.71$) for the α_{1D} -adrenoceptor subtype and weaker affinities at the α_{1A} -adrenoceptor ($pK_i = 6.23$) and α_{1B} -adrenoceptor ($pK_i = 7.86$). In comparison to its potent affinity at the α_{1D} -adrenoceptor subtype, A-315456 was 3020-, 794- and 38-fold weaker at the dopamine D_2 -, 5-HT $_{1A}$ -, and α_{2A} -adrenoceptors, respectively. These studies indicate that A-315456 is a potent and selective α_{1D} -antagonist that may serve as a useful pharmacological ligand to probe the physiological role of the α_{1D} -adrenoceptor subtype in normal and disease states. © 2001 Elsevier Science B.V. All rights reserved.

Keywords: Adrenoceptor subtype; α_{1D} -Adrenoceptor antagonist; A-315456; BMY-7378; SNAP-8719

1. Introduction

Three α_1 -adrenoceptors have been identified, cloned, and pharmacologically characterized (Bylund et al., 1994; Hieble et al., 1995). The identification of the α_{1A} -adrenoceptor specific antagonists, WB4101 ((2,6-dimethoxyphenoxyethyl)aminomethyl-1,4-benzodioxane hydrochloride) 5-methylurapidil, RS 17053 (*N*-[2-(2-cyclopropylmethoxyphenoxy)ethyl]-5-chloro- α,α -dimethyl-1*H*-indole-3-ethanamine hydrochloride), and REC 15/2739 (8-3-[4-(2-methoxyphenyl)-1-piperazinyl]-propylcarbamoyl-3-methyl-4-oxo-2-phenyl-4*H*-1-benzopyran dihydrochloride) (Ford et al., 1996; Morrow and Creese, 1986; Leonardi et al., 1997) has improved the understanding of the function of the α_{1A} -adrenoceptor subtype expressed in different mammalian tissues such as the prostate.

For the α_{1B} -adrenoceptor subtype, the alkylating agent, chloroethylclonidine, was the only α_{1B} -adrenoceptor subtype selective antagonist available (Minneman et al., 1988). How-

ever, a new quinazoline compound, L-765,314 (4-amino-2-[4-[1-(benzyloxycarbonyl)]-piperazinyl]-6,7-dimethoxyquinazoline, has recently been reported as an α_{1B} -adrenoceptor subtype selective antagonist with 10- and 100-fold more selectivity versus the α_{1D} - and α_{1A} -adrenoceptor subtypes, respectively (Patane et al., 1998). Data derived from mice that were deficient in the α_{1B} -adrenoceptor subtype suggest an important role for this subtype in blood pressure regulation (Cavalli et al., 1997).

The α_{1D} -adrenoceptor has been identified in the human bladder (Schwinn and Michelotti, 2000) and spinal cord (Smith et al., 1999), where a potential role in modulating the bothersome symptoms associated with bladder filling and storage (such as frequency, urgency, and nocturia) has recently been proposed. The most selective α_{1D} -adrenoceptor antagonist reported to date is BMY-7378 ([8-(2-[4-(2-methoxy-phenyl)-1-piperazinyl]ethyl)-8-azaspiro[4.5]decane-7,9-dione dihydrochloride]), as described by Goetz et al. (1995). However, this agent also demonstrates significant affinity for the rat recombinant 5-HT $_{1A}$ and human dopamine D_2 binding sites. More recently, Konkel et al. (1998) described SNAP-8719 (8-[(1*R*)-1-methyl-2-[4-(2,4,5-trifluorophenyl)-1-piperazinyl]ethyl]-8-Azaspiro[4.5]decane-7,9-dione hydrochloride), a structural analogue of BMY-7378, as a potent α_{1D} -adrenoceptor antagonist with reduced

* Corresponding author. Neuroscience Research, Global Pharmaceutical Research and Development, Abbott Laboratories, D-R4MN, Bldg. AP9, First Floor, 100 Abbott Park Road, Abbott Park, IL 60064-6118, USA. Tel.: +1-847-937-2694; fax: +1-847-937-9195.

E-mail address: steven.a.buckner@abbott.com (S.A. Buckner).

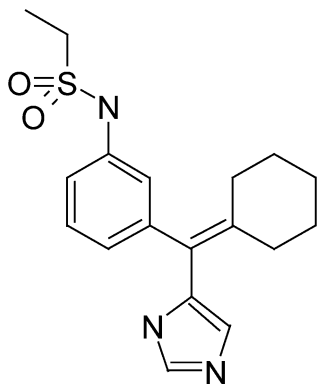


Fig. 1. Chemical structure of A-315456, *N*-[3-(cyclohexylidene-(1*H*-imidazol-4-ylmethyl))phenyl]ethanesulfonamide.

affinities at the α_{1A} - and α_{1B} -adrenoceptors, the dopamine D_2 and 5-HT $_{1A}$ binding sites. However, no functional data was presented.

The present study characterizes the α_{1D} -adrenoceptor selectivity of A-315456 (Fig. 1) using radioligand binding techniques and isolated tissue bath assays to directly compare the results to those published on the current α_{1D} -adrenoceptor subtype selective antagonists, BMY-7378, and a related analogue, SNAP-8719. The results reported here suggest that A-315456, like SNAP-8719, could serve as a useful pharmacological agent for further understanding of the role of the α_{1D} -adrenoceptor subtype *in vivo*.

2. Materials and methods

2.1. Radioligand binding assays

Studies were carried out in accordance with guidelines outlined by the Institutional Animal Care and Use Committee of Abbott Laboratories and the European Community guidelines for the use of experimental animals. Radioligand binding at the adrenergic receptor sites was performed according to Knepper et al. (1995). Briefly, the cDNA encoding bovine α_{1A} -, rat α_{1D} - and hamster α_{1B} - (Lomasney et al., 1991; Cotecchia et al., 1988) adrenoceptors were obtained from Triangle Universities Licensing Consortium (Research Triangle Park, NC), expressed in mouse fibroblast cells (LTK $^+$), and membranes were prepared and frozen at -70°C until the time of assay. Dopamine D_2 binding assays were performed using the method described by Vessotskie et al. (1997), using membranes derived from CHO cells expressing recombinant receptors (New England Nuclear, Boston, MA) and the dopamine D_2/D_3 selective agonist [^{125}I]-PIPAT under high affinity conditions. Radioligand binding assays for the rat 5-HT $_{1A}$ -receptor was performed as described by De Vry et al. (1998) using membranes derived from rat cortex expressing recombinant receptors and [^3H]-8-hydroxy-DPAT (New England Nuclear).

2.2. Functional assays

The epididymal portion of the rat vas deferens α_{1A} -adrenoceptors (Burt et al., 1992), the rat spleen α_{1B} -adrenoceptors (Han et al., 1987), the rat aorta α_{1D} -adrenoceptors (Buckner et al., 1996), and the prostatic rat vas deferens α_{2A} -adrenoceptors (Connaughton and Docherty, 1990) were employed as functional subtype specific models. Phenylephrine was used as the reference agonist for all α_1 -adrenoceptor models. Clonidine was the reference agonist for the electrical field-stimulated α_{2A} -adrenoceptor model. A-315456 was allowed a 30-min exposure time prior to generating the second agonist concentration–response curve in the presence of A-315456. Only one concentration of A-315456 was used per tissue.

2.3. Drugs and chemicals

A-315456 (*N*-[3-(cyclohexylidene-(1*H*-imidazol-4-ylmethyl))phenyl]ethanesulfonamide), BMY-7378 ([8-(2-[4-(2-methoxy-phenyl)-1-pierazinyl]ethyl)-8-azaspiro[4.5]decane-7,9-dione dihydrochloride]), SNAP-8719 (8-[(1*R*)-1-methyl-2-[4-(2,4,5-trifluorophenyl)-1-piperazinyl]ethyl]-8-azaspiro[4.5]decane-7,9-dione hydrochloride) and clonidine HCl were synthesized at Abbott Laboratories, USA. L-phenylephrine HCl and (\pm)-propranolol HCl were purchased from Sigma, St. Louis, MO, USA.

2.4. Data analysis

Concentration–response curves were analyzed using a four-parameter curve fitting routine (Zielinski and Buckner, 1998). The maximum peak amplitude response was measured and used for analysis. Data were analyzed as *g* of tension and calculated as a percentage of maximum response, and $\text{EC}_{50\text{s}}$ were determined. Antagonist potencies were expressed as pA_2 values \pm S.E.M.; slopes were not different from unity. Differences in pA_2 values were compared by analysis of variance (ANOVA), followed by Fisher's probabilistic least significant difference (PLSD) test for significance. For SNAP-8719 in the rat spleen α_{1B} -adrenoceptor assay, estimates of antagonism (pK_B) were calculated, where $K_\text{B} = [\text{antagonist, M}]/[\text{CR-1}]$ (Furchgott, 1972), and expressed as the negative $\log_{10}(\text{pK}_\text{B})$.

3. Results

3.1. Radioligand binding assays

A-315456 showed high affinity ($\text{pK}_\text{i} = 8.71$) for the α_{1D} -adrenoceptor subtype expressed in a rat clonal cell line (Table 1). In comparison to its affinity for the α_{1D} -adrenoceptor subtype, the agent was weaker at the bovine clonal α_{1A} -adrenoceptor subtype ($\text{pK}_\text{i} = 6.23$) and at the hamster clonal α_{1B} -adrenoceptor subtype ($\text{pK}_\text{i} = 7.86$). A-315456

Table 1

Radioligand binding affinities and functional potencies for A-315456, BMY-7378, and SNAP-8719

	A-315456	BMY-7378	SNAP-8719
<i>Radioligand binding, pK_i</i>			
Rat clonal α_{1D}	8.71 \pm 0.12 (1)	9.05 \pm 0.05 (1)	9.42 \pm 0.05 (1)
Bovine clonal α_{1A}	6.23 \pm 0.05 (302)	7.42 \pm 0.08 (43)	5.50 \pm 0.07 (8317)
Hamster clonal α_{1B}	7.86 \pm 0.66 (7)	7.33 \pm 0.04 (53)	7.76 \pm 0.13 (46)
Human clonal α_{2a}	7.13 \pm 0.11 (38)	5.93 \pm 0.09 (1318)	< 5.00 (>25,000)
Human clonal D_2	5.23 \pm 0.05 (3020)	7.62 \pm 0.10 (27)	5.98 \pm 0.04 (2754)
Rat clonal 5-HT $_{1A}$	5.81 \pm 0.05 (794)	8.91 \pm 0.04 (1.4)	6.47 \pm 0.02 (891)
<i>Functional antagonism, pA_2</i>			
Rat aorta α_{1D}	8.34 \pm 0.05 (1)	8.39 \pm 0.07 (1)	8.72 \pm 0.08 (1)
Rat vas deferens α_{1A}	< 4 (>10,000)	5.98 \pm 0.15 (257)	< 4.0 (>10,000)
Rat spleen α_{1B}	6.42 \pm 0.09 (83)	7.24 \pm 0.10 (14)	6.58 ^a \pm 0.16 (138)
Rat vas deferens α_{2A}	< 5 (>2000)	< 5 (>2000)	< 5 (>5000)

Data expressed as radioligand binding, pK_i (mean \pm S.E.M., $n=4-11$) and functional antagonism, pA_2 (mean \pm S.E.M., $n=8-24$). Schild slopes were not different from unity. The number in parentheses represents selectivity relative to the α_{1D} -adrenoceptor.

^a pK_B (mean \pm S.E.M., $n=12$, slope < unity).

was also examined for its affinity at 75 ancillary receptors and ion channels, and demonstrated affinity for only three additional receptors, dopamine D_2 - ($pK_i=5.23$), 5-HT $_{1A}$ - ($pK_i=5.81$), and α_2 -adrenoceptor ($pK_i=7.13$) (Table 1).

BMY-7378 showed greater affinity for all three α_1 -adrenoceptor subtypes in the current study than was reported by Goetz et al. (1995), Table 1. In our hands, the affinity (pK_i) of BMY was 9.05, 7.42, and 7.33 for the α_{1D} -, α_{1A} -, and α_{1B} -adrenoceptor subtypes, respectively. BMY-7378 showed high affinity for the dopamine D_2 -receptor, $pK_i=7.62$, even greater affinity for the 5-HT $_{1A}$ -receptor, $pK_i=8.91$, and weaker affinity for the human clonal α_{2a} -adrenoceptor, $pK_i=5.93$.

Recently, Konkel et al. (1998) presented a series of BMY-7378 analogues that identified SNAP-8719 as a potent α_{1D} -adrenoceptor subtype selective agent ($pK_i=8.89$), potency confirmed here. SNAP-8719 was tested in these same assays and generated affinities $pK_i=9.42$, 5.50, and 7.76 for the α_{1D} -, α_{1A} - and α_{1B} -adrenoceptors, respectively (Table 1); results slightly higher than those reported by Konkel et al. (1998). SNAP-8719 showed affinity for the dopamine D_2 -, 5-HT $_{1A}$ -, and the human clonal α_{2a} -adrenoceptor, with potencies (pK_i) of 5.98, 6.47, and < 5.0, respectively; results weaker than those observed at the α_{1D} -adrenoceptor.

3.2. Functional assays

In isolated tissue bath studies, A-315456 was a potent antagonist of the α_{1D} -adrenoceptor subtype ($pA_2=8.34$), as assessed by the competitive blockade of phenylephrine-induced contractions of the rat aorta (Table 1). At a maximum concentration of 100 μ M, A-315456 showed no antagonist activity at the α_{1A} - or α_{2A} -adrenoceptor examined in the phenylephrine-stimulated epididymal rat vas deferens or the field-stimulated prostatic rat vas deferens, respectively. As an antagonist of the α_{1B} -adrenoceptor subtype ($pA_2=6.42$), A-315456 was 83-fold weaker than that observed at the rat aorta

α_{1D} -adrenoceptor, as assessed in the phenylephrine-stimulated rat spleen. With the exception of the field-stimulated prostatic rat vas deferens, A-315456 showed no intrinsic stimulatory activity at any of the functional receptor sites examined up to a maximum concentration of 100 μ M. However, in the field-stimulated vas deferens assay (α_{2A} -adrenoceptor assay), A-315456, BMY-7378, and SNAP-8719 caused a concentration-dependent increase in the twitch response, thus limiting the analysis to concentrations < 100 μ M.

BMY-7378 was also a potent antagonist at the α_{1D} -adrenoceptor subtype with a $pA_2=8.39$ (Table 1). Goetz et al. (1995) reported a $pA_2=8.9$. BMY-7378 showed weak activity at the rat epididymal vas deferens α_{1A} -adrenoceptor ($pA_2=5.98$), modest potency at the rat spleen α_{1B} -adrenoceptor ($pA_2=7.24$), and no activity at the prostatic vas deferens α_{2A} -adrenoceptor ($pA_2<5$) (Table 1). The potency of SNAP-8719 was determined in these same assays and was found to be the most potent antagonist at the α_{1D} -adrenoceptor ($pA_2=8.72$), demonstrated no activity at the α_{1A} -adrenoceptor ($pK_B<4$), and demonstrated modest noncompetitive inhibition at the α_{1B} -adrenoceptor ($pK_B=6.58$) (Table 1).

4. Discussion

In radioligand binding studies, the relative affinities of A-315456 for the α_1 -adrenoceptor subtypes were $\alpha_{1D}>\alpha_{1B}$ (7-fold) $>\alpha_{1A}$ (302-fold), Table 1. A more complete biochemical study in 75 additional receptors showed that A-315456 was inactive at all receptors examined except for the dopamine D_2 -, 5-HT $_{1A}$ -, and α_{2a} -adrenoceptor where it was 3020-, 794- and 38-fold weaker, respectively, compared to the α_{1D} -adrenoceptor. BMY-7378 was equipotent at the α_{1D} -adrenoceptor and the 5-HT $_{1A}$ -receptor, and 27-fold selective against the dopamine D_2 binding site. The α_1 -adrenoceptor subtype

selectivity for BMY-7378 was $\alpha_{1D} > [\alpha_{1B} \text{ (53-fold) and } \alpha_{1A} \text{ (43-fold)}]$. Compared to its activity at the α_{1D} -adrenoceptor, SNAP-8719 showed greater than 8000-fold weaker affinity at the α_{1A} -receptor ($pK_i = 5.50$) and 46-fold weaker affinity at the α_{1B} -adrenoceptor ($pK_i = 7.76$). Radioligand binding affinities at the dopamine D_2 -receptor ($pK_i = 5.98$) and 5-HT $_{1A}$ -receptor ($pK_i = 6.47$) were less than seen with BMY-7873, but were greater than seen with A-315456 (Table 1). However, the greater α_{1D} -adrenoceptor affinity of SNAP-8719 compared to A-315456 created similar selectivity ratios. The weaker affinity of A-315456 and SNAP-8719 for the dopamine D_2 and 5HT $_{1A}$ -receptor sites makes the two compounds the most selective α_{1D} -adrenoceptor antagonists published, to date, with respect to ancillary binding activity.

α_1 -Adrenoceptor subtype selectivity was observed for A-315456 in functional tissue bath assays with $\alpha_{1D} > \alpha_{1B}$ (83-fold) $\gg [\alpha_{2A} \text{ and } \alpha_{1A} > (2000\text{- to } 10,000\text{-fold})]$. The observation that the functional antagonism at the α_{1A} -adrenoceptor subtype expressed in the rat vas deferens was less than that predicted by the radioligand binding affinity is difficult to explain. However, A-315456 (100 μ M) was also inactive at the functional α_{1A} -adrenoceptor subtype expressed in both the rabbit urethra and canine prostate gland (data not shown). Furthermore, A-315456 showed comparable radioligand binding affinity on another α_{1A} -adrenoceptor model, the rat submandibular gland with a $pK_i = 6.64 \pm 0.03$. A-315456 did not demonstrate any solubility problems, and increasing the exposure time to 60 min did not alter the data. Since A-315456 did not demonstrate any intrinsic agonist activity in the functional α_{1A} -adrenoceptor models, one could eliminate interference associated with partial agonism as a contributing factor. One might speculate a role for inverse agonism where an “agonist may behave as a positive and inverse agonist on the same receptor and differ in the stimulus pattern they produce in physiological systems” (Kenakin, 2001; Rossier et al., 1999).

BMY-7378 was also a potent antagonist at the functional α_{1D} -adrenoceptor subtype, however, it was less selective than A-315456 with $\alpha_{1D} > \alpha_{1B}$ (14-fold) $\gg \alpha_{1A}$ (>257 -fold). Although Konkel et al. (1998) did not report functional data for SNAP-8719, we were able to demonstrate potent α_{1D} -, no α_{1A} -, and only moderate α_{1B} -adrenoceptor antagonism using the same assays described above. The potency of SNAP-8719 at the spleen α_{1B} -adrenoceptor was expressed as a pK_B since the slope of the Schild plot was significantly less than unity. This may be due to a solubility issue at the higher concentrations. SNAP-8719 tends to adhere to glass, a problem solved by dissolving in dimethyl sulfoxide or methanol; however, precipitation is seen when the solution is then added to the tissue bath.

SNAP-8719 was more potent than A-315456 at the α_{1D} -adrenoceptor but also more potent at the α_{1B} -adrenoceptor subtype. The α_{1D} to α_{1B} selectivity of A-315456 and SNAP-8719 were similar, 83- and 138-fold, respectively. None of the three agents that were tested showed any functional antagonism against clonidine attenuation of field-stimulated

twitch in the rat vas deferens α_{2A} -subtype. However, they all enhanced the stimulated twitch at concentrations $>100 \mu$ M, thus limiting the analysis to concentrations lower than 100 μ M. The mechanism for this effect is not known; we have only seen this type of response when an uptake inhibitor like cocaine is given in the presence of an α_2 -adrenoceptor antagonist like rauwolscine.

The importance of the α_{1D} -subtype in the functional control of micturition has not been established, and agents like A-315456 and SNAP-8719 may play an important role in exploring this issue. It has been demonstrated that the α_{1D} -adrenoceptor present in the rat bladder is up-regulated secondary to bladder outlet obstruction (Hampel et al., 2000), a paradigm like benign prostatic hyperplasia. The α_{1D} -adrenoceptor may serve a role in irritative and filling disorders (Schwinn and Michelotti, 2000). Spinal and supraspinal α_{1D} -adrenoceptors may also modulate bladder responses (Smith et al., 1999).

In summary, A-315456 and SNAP-8719 are potent and selective antagonists at the functional α_{1D} -adrenoceptor subtype in vitro. Additionally, both agents demonstrate weaker affinity for the dopamine D_2 and 5-HT $_{1A}$ radioligand binding sites than BMY-7378. Therefore, A-315456, as well as SNAP-8719, may serve as useful pharmacological tools for further defining the role of the α_{1D} -adrenoceptor subtype in mammalian tissues in vivo.

Acknowledgements

The authors thank Renjie Chang, John Baranowski, and Chae Hee Kang for their excellent contribution in determining the radioligand binding affinities of the compounds discussed in this study.

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